Total Synthesis of Leucinostatin D

Shigeru Kuwata,* Akihiro Nakanishi, Takashi Yamada, and Toshifumi Miyazawa

Department of Chemistry, Faculty of Science, Konan University Okamoto, Higashinada-ku, Kobe 658, Japan

Abstract: Peptide antibiotic, Leucinostatin D, was synthesized by stepwise elongation method, starting from β -alanine *t*-butyl ester. Z-octapeptide ester was converted to amide derivative and finally coupled with (4S,E)-4-methylhex-2-enoyl-L-4-methylproline.

Leucinostatins are peptide antibiotics produced by *Paecilomyses* fungi. Until now, 7 congeners are isolated in nature and determined their structures; A,B,C,D,F,H and K.¹) They are active against gram-positive bacteria, yeast and fungi. They are also known as strong mycotoxins and, therefore, they called Paecilotoxins, too.²) The structural feature of these compounds are the presence of many unusual amino acids, as well as chiral unsaturated aliphatic acid as N-terminal acyl component and various chiral diamines as an amide part. High contents of leucine and α -aminoisobutyric acid (Aib) are also their characteristic feature. Recently, Aib-rich peptides have provoked a great deal of interest in their biological activities and conformations. Therefore, to examine the structure-activity relationship of these antibiotics, we have planned to synthesize leucinostatin D, a minor component of this antibiotic's family, which has relatively simple structure among them.



Fig. 1. Structure of Leucinostatin D³)

cis-L-4-Methylproline (Pro(4Me)) was synthesized by catalytic hydrogenation of L-4-methylene-proline obtained by Wittig reaction of L-4-oxoproline derived from L-4-hydroxyproline.⁴) threo-L- β -Hydroxyleucine (HyLeu) was obtained by optical resolution of racemic form which was synthesized by condensation of 2-methylpropanal with glycine and purified by recrystallization from water several times to remove the erythro compound.^{4,5}) (2S)-N¹,N¹-Dimethylpropane-1,2-diamine (DMPD) was synthesized by alkylating

dimethylamine with O-tosyl-N-Z-L-alaninol.4)

As to the synthesis of the N-acyl part of this peptide, (4S,E)-4-methylhex-2-enoyl-L-proline (Δ MHA-Pro-OH) was tried to be synthesized as a model compound. Mild Hofmann degradation⁶) and successive hydrolysis of L- β -homoisoleucyl-L-proline methyl ester afforded this compound in optically pure state.⁷) Then, (4S,E)-4-methylhex-2-enoyl-L-4-methylproline (VI) was prepared by the same synthetic route. Z-L- β -homoisoleucyl-L-4-methylproline (VI) was prepared by the same synthetic route. Z-L- β -homoisoleucyl-L-4-methylproline methyl ester (II) to Z-L- β -homoisoleucyl-L-4-methylproline methyl ester (III), (yield 88%, mp 63-65°C, [α]²⁵D -58.5° (c1.03, MeOH)). After removal of N-protecting group by catalytic hydrogenation and successive mild Hofmann degradation (MeI, KHCO3, MeOH, room temperature 2 days), (4S,E)-4-methylproline. By hydrolysis of this ester with 1M aqueous sodium hydroxide, (4S,E)-4-methylproline (Δ MHA-Pro(4Me)-OH) (VI) (yield 81%, oil, [α]²⁵D -62.2° (c 0.92, MeOH)) could be synthesized (Scheme 1.).

$$\begin{array}{c} Z-\beta-Hile + Pro(4Me)-OMe \xrightarrow{DCC-HOBt} Z-\beta-Hile-Pro(4Me)-OMe \xrightarrow{H_2 / Pd-C} \\ (I) & (II) \\ \beta-Hile-Pro(4Me)-OMe \xrightarrow{Mel / KHCO_3} \Delta MHA-Pro(4Me)-OMe \xrightarrow{NaOH} \Delta MHA-Pro(4Me)-OH \\ (IV) & (V) & (VI) \end{array}$$

TT / D. 4 C

Scheme 1. Synthetic Route for (4S,E)-4-Methylhex-2-enoyl-L-4-methylproline (VI)

For the synthesis of the C-terminal octapeptide, stepwise elongation starting from β -alanine using methyl ester as a C-terminal protecting group followed by conversion of methyl ester to amide derivative was first tried. Condensation was mainly carried out by DCC-HOBt method. However, in the case of the introduction of *threo*- β -hydroxyleucine, condensation reaction could not proceed well by DCC-HOBt method when no protection was applied to the side chain of hydroxyleucine. Therefore, Z-HyLeu-OH was derived to mixed anhydride with isobutylchloroformate and condensed with hexapeptide methyl ester. By this method, Z-heptapeptide methyl ester could be obtained in a moderate yield and elongated to Z-octapeptide methyl ester without problem. The ester, thus obtained, was tried to hydrolyze with sodium hydroxide to convert to amide derivative. However, in this case, splitting of the N-terminal protecting group and formation of urea derivative⁸ have occurred during alkaline hydrolysis. Therefore, to avoid this side reaction, C-terminal protecting group was changed as an acid-labile *t*butyl group and the same synthesis was tried again (Fig.2).⁹

In this case, Z-octapeptide acid could be obtained by usual TFA treatment of the *t*-butyl ester. Condensation of this peptide with (2S)-N¹,N¹-dimethylpropane-1,2-diamine was carried out using DCC-HOBt as a coupling reagent. After removal of the Z group of protected C-terminal octapeptide, (4S,E)-4-methylprex-2-enoyl-L-4-methylproline was condensed by DCC-HOBt method. The raw product was purified by preparative TLC (Wakogel B-5F; developing solvent, MeOH : AcOEt = 4 : 1) and leucinostatin D was obtained as a hydrochloride.





The validity of the structure was confirmed by SIMS (m/z 1119 MH⁺) and NMR (¹H, ¹³C) analyses.¹⁰) Furthermore, synthetic leucinostatin D, thus obtained, showed similar biological activities as natural one reported in the literature.^{3a})

Acknowledgments. This work was supported in part by a grant from The Science Reasearch Promotion Fund of the Japan Private School Promotion Foundation. The authors are grateful to Professor C. Rossi of Perugia University for valuable advices during this work, Tanabeseiyaku Pharmaceutical Co. Ltd. for measurements of SIMS and biological activities, and Dr. Toshio Yokoi of Kobe Gakuin University for measurement of 400MHz NMR.

References and Notes

- 1. M. G. Quaglia, S. Fanali, A. Nardi, C. Rossi, and M. Ricci, J. Chromatography, 1992, 593, 259-263.
- Y. Mikami, K. Yazawa, K. Fukushima, T. Arai, S. Udagawa, and R. A. Samson, *Mycopathologia*, 1989, 108, 195-199.
- a) C. Rossi, L. Tuttobello, M. Ricci, C. G. Casinovi, and L. Radics, J. Antibiotics, 1987, 40, 130-133.
 b) the same compound was described as leucinostatin C by another group : J. G. Stroh, K. L. Rinehart, Jr., J. C. Cook, T. Kihara, M. Suzuki, and T. Arai, J. Amer. Chem. Soc., 1986, 108, 858-859.
- 4. S. Kuwata, N. Onda, Y. Kumano, T. Yamada, and T. Miyazawa, *Peptide Chemistry 1989*, ed by N.Yanaihara, Protein Reasearch Foundation, Osaka, 1990, pp. 221-226.
- S. Kuwata, J. Tanaka, Y. Sakamoto, N. Onda, T. Yamada, and T. Miyazawa, Chem. Lett., 1989, 2031-2032.
- F. C. M. Chen and N. L. Benoiton, Can. J. Chem., 1976, 54, 3310-3311; S. Nomoto, A. Sano, and T. Shiba, Peptide Chemistry 1978, ed by N. Izumiya, Protein Reasearch Foundation, Osaka, 1979, pp. 35-40; S. Nomoto, A. Sano, and T. Shiba, Tetrahedron Lett., 1979, 521-522.
- S. Kuwata, A. Nakanishi, N. Onda, T. Yamada, and T. Miyazawa, *Peptide Chemistry 1990*, ed by Y. Shimonishi, Protein Reasearch Foundation, Osaka, 1991, pp. 109-112.
- J. A. Maclaren, Austral. J. Chem., 1958, 11, 360-365; S.G.Waley and J. Watson, J. Chem. Soc., 1953, 475-479; M. Bodanszky, J. T. Sheehan, M. A. Ondetti, and S. Lande, J. Amer. Chem. Soc., 1963, 58, 991-997; M. Waki and N. Izumiya, Bull. Chem. Soc. Jpn., 1968, 41, 1909-1916.
- Abbreviations: DCC, dicyclohexylcarbodiimide; EDC, 1-ethyl-3-(3-dimethylamino-propyl)carbodiimide; HOBt, 1-hydroxybenzotriazole; IBCF, isobutylchloroformate; MA, mixed anhydride; NMM, N-methylmorpholine; OSu, N-hydroxysuccinimide ester; TFA, trifluoroacetic acid; Z, benzyloxycarbonyl.
- 10. ¹H NMR (CDCl₃) -CH=CHCO- δ 6.36 (dd, J=1Hz, 15.4Hz, 1H); -CH=CHCO- δ 6.67 (dd, J=7.3Hz, 15.4Hz, 1H); -CO-NH- δ 8.95 (d, J=6.6Hz, 1H): δ 8.72 (d, J=4.5Hz, 1H): δ 8.35 (dd, J=3.9Hz, J=9.2Hz, 1H): δ 8.28 (s, 1H): δ 8.12 (s, 1H): δ 8.07 (d, J=5.3Hz, 1H): δ 7.70 (s, 1H): δ 7.47 (d, J=14.5Hz, 1H): δ 7.26 (d, J=6.3Hz, 1H); -N⁺H(CH₃)₂ δ 8.02 (br, 1H); -N⁺H(CH₃)₂ δ 3.16 (d, J=3.1Hz, 3H): δ 3.07 (d, J=2.8Hz, 3H).

(Received in Japan 1 July 1992)